

Synthesis of an oxa-lipoic acid[☆]

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Abstract—The synthesis of an oxa-lipoic acid is reported here for the first time. We have achieved the synthesis of the title compound starting from commercially available acrolein using a Michael addition and Knoevenagel condensation as key reactions.
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Lipoic acid (LA) **1**, a naturally occurring octanoic acid derivative, has a critical role in energy metabolism as an essential co-factor for mitochondrial α -keto-acid dehydrogenases.¹ Besides its enzymatic role, LA has been shown to act as a micronutrient with a range of antioxidant and pharmacological properties.² More importantly, LA has been used for the treatment of diabetic complications and a recent study showed that LA can be used as an anti-obesity drug.³ Despite its very attractive biological properties, it suffers from poor bioavailability. One of the reasons for its metabolic instability is β -oxidation.⁴ Previous studies on metabolism have shown that LA is subject to extensive β -oxidation and major metabolites like bisnorlipoic acid and tetrnorlipoic acids were identified.^{4c} Recently, 3-keto-lipoic acid, an intermediate resulting from β -oxidation was detected in human plasma after oral dosing in a healthy volunteer.^{4d} To address this issue, the β -methylene unit can be replaced with a group such as $-\text{CF}_2$, *N*-alkyl, or an oxygen atom. We have designed the title compound in which we replaced the β -methylene group with an oxygen atom (Fig. 1). Although, there are several reports in the literature on the synthesis of **1** and its derivatives,^{5,6} no reports on the synthesis of the oxa-lipoic acid **2** are documented. In this Letter, we disclose the first synthesis of oxa-lipoic acid **2** starting from commercially available acrolein and benzylmercaptan.

We designed the synthesis of oxa-lipoic acid **2** such that the dithiolane moiety was introduced at a later stage due

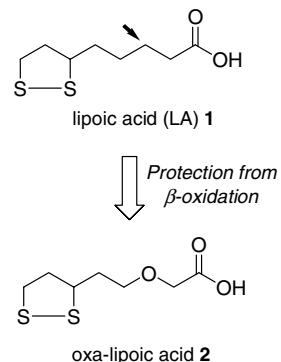


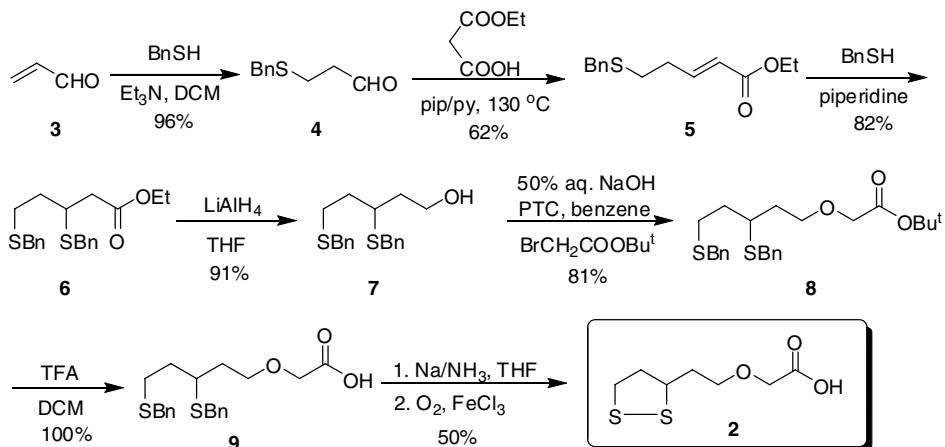
Figure 1. Protection from β -oxidation: replacement of β -methylene group with an oxygen atom.

to its sensitivity towards various reaction conditions; in particular, it is prone to oxidation. Our synthesis began with the reaction of acrolein **3** with benzylmercaptan to give the aldehyde **4**.⁷ Two-carbon homologation of compound **4** was achieved by heating with malonic acid monoethyl ester in the presence of piperidine and pyridine mixture to furnish **5** in 62% yield.⁷ Addition of another molecule of benzylmercaptan in a Michael fashion resulted in compound **6**.⁷ The alcohol **7**, obtained by LiAlH_4 reduction of **6** was subjected to O-alkylation with *t*-butyl bromoacetate under phase-transfer catalysis to furnish the ether **8** in 81% isolated yield. Removal of the *t*-butyl group in **8** by stirring with TFA in DCM gave the corresponding acid **9** in quantitative yield. The synthesis of oxa-lipoic acid **2** was achieved by construction of the dithiolane ring using a known procedure.^{6h,i} Reductive removal of the benzyl groups with sodium–ammonia followed by oxidation furnished the oxa-lipoic acid as a viscous material in 50% isolated

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**Scheme 1.**

yield over two steps. The spectral data (IR, ¹H NMR, ¹³C NMR and MS) of the **2** were in agreement with structure **2**⁸ (see Scheme 1).

In conclusion, the first synthesis of oxa-lipoic acid **2** was achieved starting from acrolein and benzylmercaptan. Compound **2** was designed to resist from β-oxidation, which may help in improving the bioavailability of lipoic acid **1** by retaining its biological activity.

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References and notes

- (a) Randle, P. J. *Diabetes Metab. Rev.* **1998**, *14*, 263; (b) Reed, L. J.; Gunsalus, I. C.; De Busk, B. G.; Hornberger, C. S., Jr. *Science* **1951**, *114*, 93; (c) Schmidt, U.; Grafen, P.; Altland, K.; Goedde, H. W. *Adv. Enzymol.* **1969**, *32*, 423; (d) Sigel, H. *Angew. Chem., Int. Ed. Engl.* **1982**, *21*, 389.
- (a) Evans, J. L.; Goldfine, I. D. *Diabetes Technol. Ther.* **2000**, *2*, 401; (b) Biewenga, G. P.; Haenen, G. R.; Bast, A. A. *Gen. Pharmacol.* **1997**, *29*, 315; (c) Packer, L.; Witt, E. H.; Tritschler, H. J. *Free Radical Biol. Med.* **1995**, *19*, 227; (d) Han, D.; Handleman, G.; Marcocci, L.; Sen, C. K.; Roy, S.; Kobuchi, H.; Tritschler, H. J.; Flohé, L.; Packer, L. *Bio Factors* **1997**, *6*, 321.
- (a) Betteridge, D. J. *Metabolism* **2000**, *49*, 3; (b) Baynes, J. W.; Thorpe, S. R. *Diabetes* **1999**, *48*, 1; (c) Paolissio, G.; Giugliano, D. *Diabetologia* **1996**, *39*, 357; (d) Wollheim, C. *Diabetologia* **2000**, *43*, 265; (e) Kim, M.-S.; Park, J.-Y.; Namkoong, C. *Nat. Med.* **2004**, *10*, 727.
- (a) Schpke, H.; Hempel, R.; Peter, G.; Hermann, R.; Wessel, K.; Engel, J.; Kronbach, T. *Drug Metab. Dispos.* **2001**, *29*, 855; (b) Harrison, E. H.; McCormick, D. B. *Arch. Biochem. Biophys.* **1974**, *160*, 514; (c) Furr, C. H.; Chang, H. H.; McCormick, D. B. *Arch. Biochem. Biophys.* **1978**, *185*, 576; (d) Biewenga, G. P.; Vriesman, M. F.; Haenen, G. R. M. M.; Bast, A. In *Lipoic Acid, A Pharmacochemical Study*; Biewenga, G. P., Ed.; Academisch Proefschrift, Vrije Universiteit: Amsterdam, 1997; pp 137–151.
- (a) Reed, L. J.; Ching, I. N. *J. Am. Chem. Soc.* **1954**, *77*, 416; (b) Page, P. C. B.; Rayner, C. M.; Sutherland, I. O. *J. Chem. Soc., Chem. Commun.* **1986**, 1408; (c) Rao, A. V. R.; Mysorekar, S. V.; Gurjar, M. K.; Yadav, J. S. *Tetrahedron Lett.* **1987**, *28*, 2183; (d) Brookes, M. H.; Golding, B. T.; Hudson, A. T. *J. Chem. Soc., Perkin Trans. 1* **1988**, *9*; (e) Laxmi, Y. R. S.; Iyengar, D. S. *Synthesis* **1996**, 594; (f) Adger, B.; Bes, M. T.; Grogan, G.; McCauley, R.; Pedragosa, M. S.; Roberts, S. M.; Villa, R.; Wan, P. W. H.; Willetts, A. J. *Bioorg. Med. Chem.* **1997**, *5*, 253; (g) Upadhyay, T. T.; Nikalje, M. D.; Sudalai, A. *Tetrahedron Lett.* **2001**, *42*, 4891; (h) Chavan, S. P.; Praveen, Ch.; Ramakrishna, G.; Kalkote, U. R. *Tetrahedron Lett.* **2005**, *45*, 6027, and references cited therein.
- (a) Koufaki, M.; Calogeropoulou, T.; Detsi, A.; Roditis, A.; Kourounakis, A.; Papazafiri, P.; Tsiaikitzis, K.; Gaitanaki, C.; Beis, I.; Kourounakis, P. *J. Med. Chem.* **2001**, *44*, 4300; (b) Koufaki, M.; Detsi, A.; Theodorou, E.; Kiziridi, C.; Calogeropoulou, T.; Vassilopoulos, A.; Kourounakis, A. P.; Rekka, E.; Kourounakis, P. N.; Gaitanaki, C. *Bioorg. Med. Chem.* **2004**, *12*, 4835; (c) Gruzman, A.; Hidmi, A.; Katzhendler, J.; Haj-Yehie, A.; Sassan, S. *Bioorg. Med. Chem.* **2004**, *12*, 1183; (d) Morera, E.; Luente, G.; Ortar, G.; Nalli, M.; Mazza, F.; Gavuzzo, E.; Spisanic, S. *Bioorg. Med. Chem.* **2002**, *10*, 147; (e) Huwe, C. M.; Kunzer, H. *Tetrahedron Lett.* **1999**, *40*, 683; (f) Harnett, J. J.; Auguet, M.; Viessat, I.; Dolo, C.; Bigg, D.; Chabrier, P. E. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1439; (g) Matsugo, S.; Yan, L. J.; Konishi, T.; Youn, H. H.; Lodge, J. K.; Ulrich, H.; Packer, L. *Biochem. Biophys. Res. Commun.* **1997**, *26*, 819; (h) Thomas, R. C.; Reed, L. J. *J. Am. Chem. Soc.* **1956**, *78*, 6151; (i) Thomas, R. C.; Reed, L. J. *J. Am. Chem. Soc.* **1956**, *78*, 6151.
- (a) Uneme, H.; Mitsudera, H.; Kamikado, T.; Kono, Y.; Manabe, Y.; Numata, M. *Biosci., Biotechnol., Biochem.* **1992**, *56*, 2023; (b) Wladislaw, B. *Chem. Ind. (London)* **1957**, 263.
- Spectral data of **2**: IR (Neat): 1731 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.54 (s, 1H), 3.99 (s, 2H), 3.77–3.71 (m, 1H), 3.54 (t, 2H, *J* = 6.4 Hz), 3.2–3.09 (m, 2H), 2.47–2.39 (m, 1H), 1.99–1.88 (m, 2H), 1.82–1.74 (m, 1H); ¹³C NMR (100 MHz, DMSO): δ 171.6, 69.4, 67.4, 52.8, 40.7, 39.8, 38.2; MS (Cl): 208 (M⁺).